

Survey Plan for Adult Fish Counting and Coho Prespawn Mortality in Miller and Walker Creeks

Community Salmon Investigation for Highline 2010

Version October 27, 2010

NOAA, Wild Fish Conservancy, the City of Seattle, King County and others have conducted a number of assessments concerning coho salmon in Seattle area creeks (Collier et al. 2003, Scholz et al. 2004, and McCarthy et al. 2008). Findings of these efforts indicate that while salmon were successfully returning to many urban streams, a high proportion of sexually mature female coho carcasses showed large numbers of retained eggs. Investigators documented highly erratic swimming behavior and prespawn mortality among both male and female coho. Affected fish from different urban streams displayed a common suite of symptoms, including surface swimming and gaping, fin splaying, spasming, disorientation, and loss of equilibrium. The coho usually died within a few minutes to a few hours after becoming overtly symptomatic. Visual inspections generally indicated that the affected coho spawners were in good condition, with the silver coloration typical of salmonids that have recently transitioned to freshwater from the ocean (McCarthy et al. 2008). This phenomenon has been termed coho prespawn mortality (PSM).

In Miller and Walker Creeks, PSM has been documented and is probably a serious concern for the recovery of coho populations in the basin. However, the information gathered in the Miller/Walker creek basin has been largely anecdotal, as systematic surveys of PSM have not been conducted. Yearly systematic surveys of PSM in the basin of a data quality sufficient to determine percent survival and percent PSM of adult coho returning to spawn will provide information for several purposes. The most important information will be to determine to what extent PSM occurs in the Miller/Walker basin. Severity and variability from year to year can provide clues as to causal factors. Because PSM is an ongoing phenomenon in Miller and Walker Creeks, monitoring every year for an extended timeframe could give us an indication if changes designed to improve the aquatic ecology of these creeks are having their intended effects. It may also be possible to see the degree of change of a given basin improvement based on PSM data. Collecting yearly PSM data could therefore be an important indicator of the effectiveness of our efforts to improve basin hydrology and water quality over time.

Currently, systematic surveys have only been done on Longfellow Creek in Seattle. PSM data on Miller/Walker will provide important regional data that is currently lacking and may also attract other research on the phenomena as part of a synergism of study.

In addition to gathering data on PSM, the Miller and Walker Creeks survey will systematically survey selected locations to generate the minimum estimate of adult coho and chum adult returns to the streams. A better understanding the range of adult fish returning to the streams was one of the top recommendations the community identified as part of coordinated monitoring of the

streams (<http://www.kingcounty.gov/environment/watersheds/central-puget-sound/miller-walker-creeks/monitoring.aspx>).

PSM studies and adult fish counts will be carried out by basin volunteers and organized by the basin steward in 2010. Surveys will be conducted daily during spawning season (mid-October – mid December) to collect information on the number of returning coho, including the number of returning fish, the number of PSM fish, and the number that appeared to die due to predation. Survey volunteers will collect the following measurements for each dead fish encountered: fork length, girth, and postorbital and hypural. In addition, information will be recorded concerning adipose fin presence, sex, percent egg retention, and spawning status.

The survey reaches on Miller and Walker Creeks are a combination of the area of potential spawning and the area that a survey crew can survey in about two hours each day. During 2010, there are two survey areas on each creek.

Daily stream surveys began October 8, 2010. They will conclude in mid-December.

Methods

Objective: Collect as much information as possible on living adult salmon and dead coho and chum within the stream or found on land. Spawning status is more important for females than males. If you can determine male status, record it, otherwise record as unknown.

Day of Sampling:

Be sure to take each of the items below to the field:

Sampling kits

- data sheets, site maps and clipboard (you provide clipboard)
- pencils for recording data
- Sharpie pen for marking “flags”
- folding knife
- orange Zak knife for gutting fish
- tape measure
- flagging tape
- gloves (work gloves for handling fish)
- paper towels
- garbage bags (to carry out all trash)

Video Camera (optional) – can use video on digital camera if necessary (for documenting symptomatic fish,)

Rain gear, boots, waders

Stream Sampling Instructions

If it has rained and the creek is cloudy or if the water level is high you will probably not be able to see fish or redds. It may also be impossible to walk in the stream and dangerous to walk along it. Do not attempt to walk the stream. Wait until visibility is better (this may require cancelling the survey in part or totally for a given day).

If the creek is clear, walk the stream to see if there are live symptomatic fish and/or mortalities (videotape symptomatic fish if equipped to do so).

Watch for redds, mark the redds (see “Flagging Redds” below) and don’t step on the redds!

Further information on how to walk the streams safely and observe fish are found in the separate document titled “How to Survey for Adult Salmon.”

Recording Data on the Daily Survey Sheet

The formatting of the Daily Survey Sheet has been modified several times at the beginning of the 2010 survey season to improve its ease of use by volunteers. The most up-to-date version of the survey form always will be posted at the “CSI: Highline” web page:

<http://www.kingcounty.gov/environment/watersheds/central-puget-sound/miller-walker-creeks/salmon-monitoring.aspx>

The Daily Survey Sheet: TOP

- Fill in **DATE**. This means there is a new worksheet for every day of sampling.
- **SURVEYORS’ NAMES**: First initial and last name of all surveyors.

The Daily Survey Sheet: BOTTOM

- Fill in **START TIME** and **END TIME**.
- Fill in **WEATHER**. Check more than one if applicable.

The Daily Survey Sheet: Dead Adult Fish

- **FISH ID# is the label you will assign to each fish recorded (see below for formula)**
- **FORK LENGTH**: measure in centimeters. This is the measure from the tip of the nose to the **indent (fork) of the tail**.
- **GIRTH**: distance around largest section of fish (typically in front of dorsal fin)

Important Point: PSM investigation is for ADULT fish only. If you encounter dead juvenile fish, describe them on your form but do not conduct necropsy.

- **POH:** distance from postorbital (behind eye) to hypural plate (point in tail where it will not bend upward, last vertebrae are fused to support caudal rays)
- **ADIPOSE FIN PRESENT?** Y = Yes, N = No.
- **SEX:** M = Male, F = Female, UNK = Unknown
- **% EGG RETENTION:** Choose from 0-25, 25-50, 50-75 or 75-100%.
- **SPAWNING CONDITION/PREDATION:** Mark either:
 - Pre-spawn (PSM) for females full of eggs w/ no sign of predation. Also check the PSM box if you find a male with clearly full testes.
 - Post-spawn (POST) for females and males that are spawned out and have no sign of predation.
 - Predation (PRED) for males or females if they have been eaten in any way.
 - Unknown (UNK) for fish where pre- or post- spawning condition is undeterminable.
- **SPECIES:** Identify species: **coho**, **chum**, or **unknown**
- **NOTES:**
 - Write notes on anything you observe about the fish that is unusual. For example, if there is evidence of predation, how much of the fish has been eaten? Were parts missing?



NEW



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IMPORTANT

Important Reminder: Only count tails to avoid double counting fish so, if you find only a head or fish parts, do not count it as a dead fish found that day. On the other hand, if you only find a tail, punch/slit a hole in it, flag it with a **PRED** + *the date* label on the flagging tape and count it as a mortality for the day.

FISH ID#

Each fish mortality will receive a Fish ID#. The Fish ID # follows the formula:

Date+Site+Status+sequential number

EXAMPLE 1: *101510UprMlr_PSM01* is first dead fish found on Oct 15th at Upper Miller Creek survey site, and it was a pre-spawn mortality.

EXAMPLE 2: *101710UprWlr_UNK01* is first dead fish found on Oct 17th at Upper Walker Creek survey site, and it was too heavily predated to determine whether it had spawned or not.

Use acronyms for fish condition as defined on the field sheet (PSM, POST, PRED or UNK). Writing in pencil will allow you to modify the coding if you change your assessment of likely mortality during the necropsy.



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IMPORTANT

Flagging Fish Mortalities

Carefully cut a slit/hole in the meaty part of the tail (using the folding knife) of any fish that you count so that following groups will not count the same fish again. Cut a long piece of flagging

tape and mark “2010 PSM STUDY + *the date*” on it. Then tie the flagging tape through the hole you made in the fish tail. The tape should be large enough so it can be seen easily underwater. Return fish to where you found it in the wetted area of the stream. Whittle a stake from a tree branch and drive it into the body behind the gill cover and into the stream substrate. If the carcass is out of the water, put it back in. Anchoring will lengthen the time the carcass stays in the stream so it is more available for predation by aquatic organisms and decomposition.



Sexing Fish

Determine the sex of fish by making an incision using the orange Zak knife on the ventral surface of the body from a point immediately anterior to the anus toward the head to a point immediately posterior to the pelvic fins. If necessary, a second incision should be made on the other side of the fish from the initial point of the first incision toward the dorsal fin. The resulting flap can be folded back to observe the gonads. Ovaries have a granular texture and depending on the species can range from orange/red to dark green/blue or even whitish in color. Testes appear creamy off-white and have a smooth texture. Record the sex of each fish on the Processing Bench Sheet using M for male, F for female, U for unknown. Add a question mark (?) after M or F for unsure. Fish will be categorized in one of the following 4 categories:

Pre-spawn mortality (PSM) = fish that are dead and have not spawned completely.

We will collect the following information from pre-spawn mortalities:

- Assign a FISH ID# (see “FISH ID#” above). You may wish to wait to assign the complete fish number when you have completed the necropsy and can determine with confidence what the condition of the fish is.
- Fork Length
- Girth
- Post-orbital to hypural length (behind the eyeball to the “flex” in the tail)
- Percent egg retention for females
- Presence/absence of adipose fin



Post-spawn mortality (POST) = fish that are dead and have spawned completely. Collect the same information from post-spawners as from PSM fish.

Preyed upon mortality (PRED) = fish that are dead and have been preyed upon. Collect as much of the same information as possible from the preyed upon fish as from PSM fish. **Be sure to write “PRED” in the “note section” on Daily Survey Sheet.**

Unknown (UNK) = fish for which pre- or post-spawn status can’t be determined (that is, if predation or damage is too severe).

The Daily Survey Sheet: Live Adult Fish

Record the number of live adult fish seen in each survey area. Record as “coho” or “chum” those fish that you are confident you have identified correctly. You should be able to see at least two distinctive features of each species to be able to ID the fish as a given species. If in doubt about the species, DO NOT GUESS. Instead, record it as an “Other Adult Fish.”

Use the “Notes” section for each survey area to record anything of interest including:

- Adult symptomatic fish = adults opening and closing mouth rapidly (gaping), loss of equilibrium, fins splayed, and spasming. These behaviors are correlated with pre-spawn mortality.
- Juvenile fish observed
- Wildlife observed
- Stream conditions including clarity of water, flow volume, presence/absence of foam

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The Daily Survey Sheet: Suspected Redds

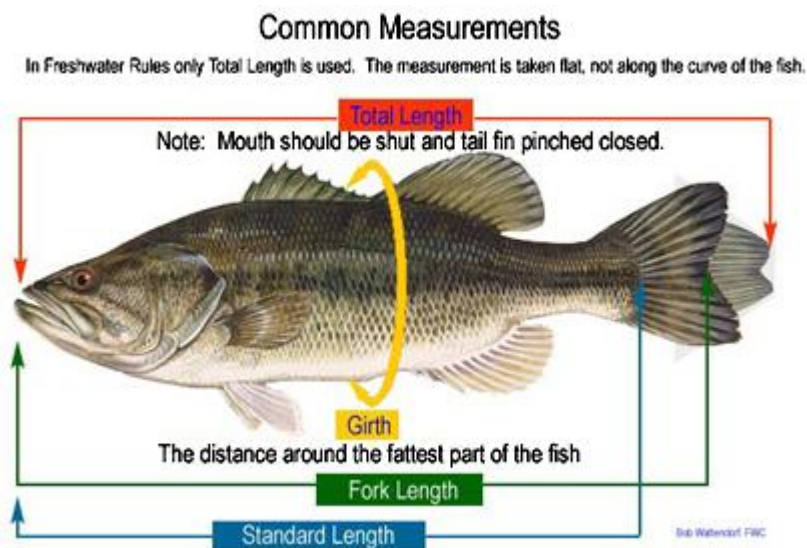
Use this line to make notes about any redds identified as new and marked for the first time.

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Flagging Redds

Flag any new redds with flagging tape. Write “REDD PSM STUDY + the date” on the flagging tape and tie it on a branch above the redd so that samplers will not walk in the redd on future surveys. If there is no convenient branch overhead, find a large rock to tie flagging to and drop in the pit of the redd, not on the mound. (All flagging from both carcasses and redds will be removed by the final survey crew in December.)

How to Measure Freshwater Fish (from Florida Fish and Wildlife website)



Fork Length Measurement

The **fork length** is the length of the fish to the fork in the tail, with the mouth closed.

1. Lay fish flat



2. Pinch mouth shut and align with front of tape

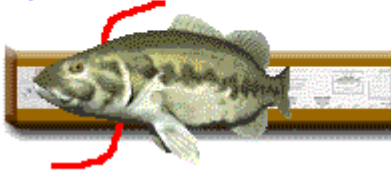


Girth Measurement

"Girth" is best measured with a fabric ruler, such as tailors use. It can also be determined by drawing a string around the fish at its widest point marking where the string overlaps and then measuring the distance between the overlapping points on a conventional ruler. The measurement should be taken perpendicular to the length of the fish. This measurement is analogous to measuring the circumference of someone's waist.

Knowing the girth is important when trying to certify a fish for a record, and provides useful information to biologists about the relative condition of a fish.

1. Gently lift fish up and slide a piece of fishing line or a flexible tape measure under fish.

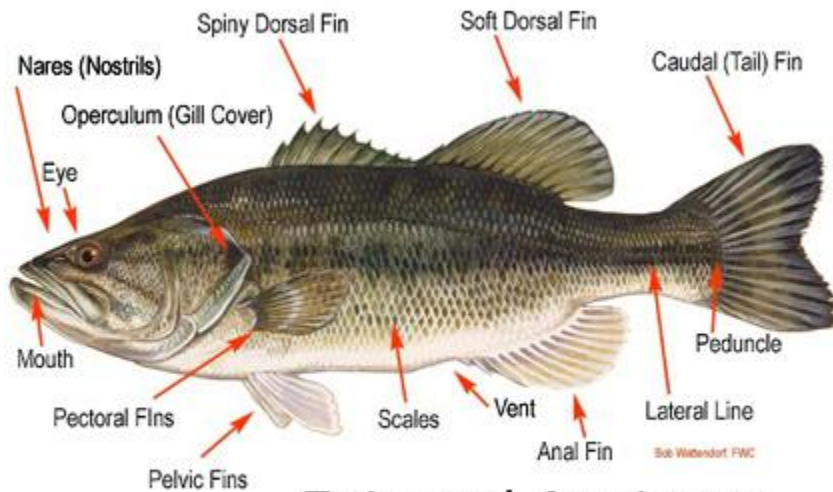


2. Lay fish flat with line or tape under deepest part of fish. Wrap it around, fold fins down if needed, line should be perpendicular.



Mark where line crosses.

3. Gently release fish. Remember minimize the fish's time out of water. Lay marked line on tape measure, pull tight, and read girth.



External Anatomy

Internal Fish Anatomy

The following illustration of a largemouth bass shows some of the common internal features that are used to describe the differences between fish that are described in more detail below.

